

Drug Design Progress of In silico, In vitro and In vivo Researches

Qifeng Bai^{1,*,#}, Lanlan Li^{2,*,#}, Shanhui Liu², Shujun Xiao^{1,2}, Yu Guo^{1,2}

¹Key Lab of Preclinical Study for New Drugs of Gansu Province, School of Basic Medical Sciences, Lanzhou University, Lanzhou, Gansu 730000, P. R. China

²Institute of Urology, Lanzhou University Second Hospital; Key Laboratory of Urological Diseases in Gansu Province, Lanzhou University; Gansu Nephro-Urological Clinical Center, Lanzhou, Gansu, China, 730030

#These authors contributed equally to this work.

Abstract

Drug design, referred to the fields of pharmacology, biotechnology and medicine, is *in silico*, *in vitro* and *in vivo* assay processes of finding new candidate medications based on the biological targets. The *in silico* experiments of drug discovery are involved in the macromolecular structure databases, small molecule databases, molecular docking, de novo drug design and molecular dynamics simulations. The *in vitro* experiments of drug discovery need evaluate the direct interaction information between ligands and targets as well as the function of ligands on signaling pathway in the cell. The *in vivo* experiments of drug discovery give the convincing evidence for preclinical trial at the physiological level. In this review, we outline the drug design components of databases, virtual screening tools, biochemical assays, cell-based system and animal models.

Corresponding Author: Qifeng Bai, Key Lab of Preclinical Study for New Drugs of Gansu Province, School of Basic Medical Sciences, Lanzhou University, Lanzhou, Gansu 730000, P. R. China, Email: baifq@lzu.edu.cn; Lanlan Li, Institute of Urology, Lanzhou University Second Hospital; Key Laboratory of Urological Diseases in Gansu Province, Lanzhou University; Gansu Nephro-Urological Clinical Center, Lanzhou, Gansu, China, 730030, Email: llli12@lzu.edu.cn

Keywords: virtual screening tools, biochemical assays, cell-based system and animal models

Received: May 05 , 2018

Accepted: Aug 03, 2018

Published: Augs 16, 2018

Editor: George Kordas, Twice ERC laureate (Nanotherapy Advanced Grant and PoC Grant) ERC LS7 Panel Member, Member of the ASF 15 Panel

Introduction

Modern new drug design is the integrated and long-term processes which will cost tens to hundreds of millions dollars from candidate compounds trial to Food and Drug Administration (FDA) approval¹. The preclinical drug design trial can be made of *in silico*, *in vitro*, and *in vivo* experiments. The development of information technology and big data accelerate the speed of drug discovery because the high effective and targeted databases are constructed under these circumstances². The famous databases designed for drug discovery are introduced in detail following review part. Based on the accurate structural models of biological targets and small molecules, the molecular docking software³ can be used to estimate the affinity of ligand in the pocket of biological macromolecules. In addition, the de novo method is another computer-aided design for new drug generation on basis of 3D-structural targets and pharmacophore model⁴. To study the dynamical interaction between receptors and ligands at the atomic level, the molecular dynamics (MD) simulations supply a reliable and accurate way to explore the binding mechanism between ligands and targets^{5,6}. Our review describes the popular molecular docking, de novo drug and MD simulation software in drug discovery field. The candidate compounds screened *in silico* still need be validated to make sure the compound has pharmaceutical activity. The direct binding experiment *in vitro* between ligands and targets should be trial by using the methods of X-ray crystallography structural analysis, Surface Plasmon Resonance (SPR), etc⁷. Because the direct binding assay cannot guarantee the activity of screened ligands, the cell signaling pathway response experiment should be performed to check the activity of ligands in cell⁸. Due to the complex physiological environment, the active ligands *in vitro* trial may not show any response to the targeting disease *in vivo*. Hence, it need choose the suitable animal models for *in vivo* experiments⁹. In this review, we introduce the database, *in silico* drug design software, *in vitro* experiment, and *in vivo* animal models (see Figure 1). Of course, the sequence of *in silico*, *in vitro* and *in vivo* experiments can be changed according to specific conditions. For instance, if the animal models are easier to be got and cheaper to be bought than biochemical assays, the *in vivo* experiments can be placed before the

in vitro trial. Generally, our review gives the researchers an easy understanding contour for drug design.

In silico Database for Drug Design

With the development of bioinformatics, big data, biology, chemistry and medicine, more and more databases are design to service for the drug discovery¹⁰. The databases are divided into macromolecular and small molecular databases. The macromolecular structure databases contain the crystal structures of proteins, nucleic acids, or other biopolymers. Table 1 shows the popular databases of macromolecular crystal structures and theoretical 3D macromolecular structures by homology modeling method. The wwPDB¹¹ currently contains three Protein Data Bank (PDB) databases and one Biological Magnetic Resonance Data Bank which are RCSB PDB, PDBe, PDBj, and BMRB, respectively. The RCSB PDB database is a three dimensional (3D) structural crystallographic database for large biological molecules such as proteins and nucleic acids¹² which are collected from NMR spectroscopy, X-ray crystallography, and cryo-electron microscopy^{13, 14}. The databases of PDBe and PDBj lay in Europe and Japan are another two organizations which are responsible for the collection and dissemination of biological macromolecular structures. The Biological Magnetic Resonance Data Bank (BMRB) focuses on the data collection of NMR Spectroscopy from peptides, proteins, nucleic acids, and other biomolecules. Nucleic Acid Database (NDB)¹⁵ recruits the functions, structures, analysis, and sequences of experimentally-determined nucleic acids. Molecular Modeling Database (MMDB)¹⁶ collects experimentally resolved three-dimensional biomolecule structures under the maintenance of National Center for Biotechnology Information. JenaLib database¹⁷ emphasizes the visualization and analysis of three-dimensional biopolymer structures. PDBbind database^{18, 19} is interested in collecting the experimental binding affinity data. The molecular docking scores can be developed on basis of the collected data of Kd, Ki, and IC50 in PDBbind database. Generally, these macromolecular databases accelerate the drug discovery via providing accuracy and rich structural information, especially, the crystal target data in complex with ligands. Based on the accuracy crystal information, the successful drug screenings and mechanism studies are reported on the targets of transcription factor²⁰,

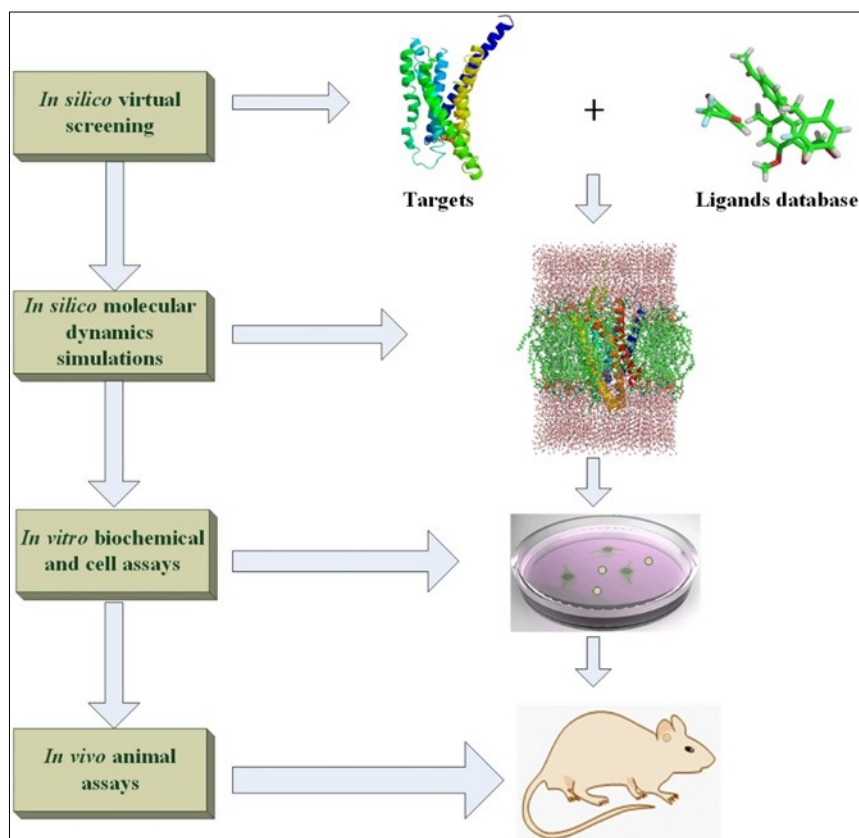


Figure 1. The diagram of *in silico*, *in vitro* and *in vivo* for drug design.

Table 1. Macromolecular structure databases

Structure databases	Description	Weblink
wwPDB	Worldwide macromolecular structures	www.wwpdb.org
RCSB PDB	Macromolecular structures	www.rcsb.org
PDBe	Macromolecular structures in Europe	www.ebi.ac.uk/pdbe
PDBj	Macromolecular structures in Japan	pd bj.org
BMRB	Macromolecular structures from NMR	www.bmrb.wisc.edu
NDB	Nucleic acid structure database	ndbserver.rutgers.edu
MMDB	3D macromolecular structures	www.ncbi.nlm.nih.gov/Structure/MMDB/mmdb.shtml
JenaLib	3D biopolymer structures	jenalib.leibniz-fli.de
PDBbind	3D macromolecular structures in complex with ligands	www.pdbbind.org.cn
IUPHAR/BPS Guide to PHARMACOLOGY	Pharmacological targets and substances	www.guidetopharmacology.org
ModBase	Theoretical 3D macromolecular structures	modbase.compbio.ucsf.edu/modbase-cgi/index.cgi
PMP	Theoretical 3D macromolecular structures	www.proteinmodelportal.org
SWISS-MODEL	Theoretical 3D macromolecular structures	swissmodel.expasy.org/repository

G protein-coupled receptors (GPCRs)²¹⁻²³, transporter²⁴, ion channels receptors²⁵, and so on. IUPHAR/BPS Guide to PHARMACOLOGY^{26, 27} provides the information of clinical, approved drugs and candidate compounds for the popular targets such as GPCRs, ion channels receptors, kinases, transporters, and so on. It is very simple for users to find the drug comprehensive resources which contain 3D crystal structure targets, relative database links, bioactive ligands from literature, antibodies, functional assays, physiological functions, disease models and so on.

Until now, the number of proteins with crystal structures is limited because there are still some intractably unresolved crystal proteins and nucleic acids structures. The targets without the crystal structures cannot supply the accurate 3D models for drug discovery. In order to screen the candidate compounds based on the active sites of targets without crystal structure, it need predict the 3D structures from the sequence of proteins or nucleic acids. The ModBase²⁸, PMP²⁹, and SWISS-MODEL³⁰ can help the researchers build the 3D theoretical models of targets by comparative modeling methods. The theoretical 3D macromolecular structures also give the reliable 3D models to screen the potential drugs from the large of small molecule database. Some studies have shown homology model can be considered as the valid target to perform the virtual screening^{31, 32}. The crystal and predicted models have been widely applied into the research field of drug discovery.

Drug discovery not only needs the reliable models, but also is relied on huge of small molecules with reasonable conformations. Table 2 shows the popular and special purpose small molecule databases for drug discovery. ZINC database^{33, 34} is considered as an open-access commercially-available screening library which contains over 35 million purchasable small molecules for virtual screening. ZINC database, which supplies Lead-Like, Drug-Like, Fragment-Like for different research needs, has been reported to be used to screen the candidate compounds by molecular docking methods^{35, 36}. PubChem database currently consists of compounds, substances and bioassay databases which recruit the 93.9 million, 249 million and 1.25 million entries, respectively. Therefore, PubChem³⁷

database can be used to dig out the potential bio-activated compounds through molecular docking and deep learning studies. DrugBank³⁸ is a comprehensive and open-access database containing detail information of drug data and drug targets. In combination with the ZINC database, it has been used to find the ligands via molecular docking based screening^{39, 40}. The databases of ChemBridge, Specs and ChemDiv can supply with the commercially customized screening libraries for the drug development targeting to research receptors such as cyclophilin D⁴¹, *p*-Hydroxyphenylpyruvate dioxygenase (HPPD)⁴², SETDB1/ESET⁴³, and so on. The e-Drug3D⁴⁴ database recruits the 1852 molecular structures which approved by Food and Drug Administration (FDA) between 1939 and 2017 with a molecular weight \leq 2000. The e-Drug3D can be used as the benchmark for finding the effective candidate compounds. Super Natural II⁴⁵ is a database of natural compounds with physicochemical properties, predicted toxicity information, 2d structures, and vendors. In all, \sim 75% of FDA approved small molecular drugs are from natural compounds or its derivatives⁴⁶. Hence, Super Natural II database has a good promising for drug development by *in silico* method. The Cambridge Structural Database (CSD)⁴⁷ and Crystallography Open Database (COD)⁴⁸ are mainly interested in the collection of small molecule organic and inorganic crystal structures. It cannot only be used for drug virtual screening, but also give the accurate structural information for other computing such as quantum chemical calculation. The KEGG Ligand database⁴⁹ comprises COMPOUND, REACTION, and ENZYME which is responsible for collecting the chemical compounds, chemical reactions, and enzyme molecules, respectively. The nucleic acid ligand database (NALDB)⁵⁰ and small molecule pathway database (SMPDB)⁵¹ are designed for the special purpose of drug discovery. The NALDB provides the detail experimental data of small molecules which target to nucleic acid structures. The SMPDB gives the ligands interactive network on signal pathway found in human which is used to elucidate the drug-action signaling pathways. The MarinChem3D (mc3d.qnlm.ac), which locates at National Laboratory for Marine Science and Technology (Qingdao) in China, publishes more than 30,000 well-defined 3D structures of marine natural products. It can be used to perform the virtual screening studies directly. The MarinChem3D

Table 2. Small molecule databases

Ligand databases	Description	Weblink
ZINC	Over 35 million purchasable compounds for virtual screening	zinc.docking.org
PubChem	Over 90 million compounds	pubchem.ncbi.nlm.nih.gov
DrugBank	More than 11,000 drug entries	www.drugbank.ca
Specs	Providing high throughput screening compounds	www.specs.net
ChemBridge	Over 1.1 million druglike and leadlike compounds	www.chembridge.com/ screening_libraries
ChemDiv	Over 1,5 M individual solid screening compounds	www.chemdiv.com/ services-menu/screening-libraries
e-Drug3D	1852 FDA approved drugs between 1939 and 2017	chemoinfo.ipmc.cnrs.fr/ MOLDB/index.html
Super Natural II	325,508 natural compounds	bioinf-applied.charite.de/ supernatural_new/index.php
CSD	Over 900,000 small-molecule organic crystal structures	www.ccdc.cam.ac.uk/solutions/ csd-system/components/csd
COD	Over 390,000 inorganic crystals and small organic compounds	www.crystallography.net/cod
KEGG Ligand	Universe of chemical substances and reactions	www.genome.jp/kegg/ligand.html
NALDB	Nucleic acid ligand database	bsbe.iiti.ac.in/bsbe/naldb/ HOME.php
SMPDB	Over 30,000 small molecule pathways found in humans	smpdb.ca
MarinChem3D	Over 30,000 kinds of marine compounds	mc3d.qnlm.ac

gives a promising way to find candidate ligands targeting to receptors from the ocean.

In silico Software for Drug Design

Although the macromolecular structure and small molecule databases are an important factor for drug discovery *in silico*, it still needs the effective software for performing virtual screening on targets and small molecule databases. In the past decades, various molecular docking software emerges based on different algorithms and molecular formats. Generally, the computational methods for drug discovery can be divided into ligand-based (indirect) and structure-based (direct) techniques⁵². The ligand-based drug design methods contain quantitative structure-activity relationship (QSAR)⁵³, pharmacophore⁵⁴, etc. The structure-based drug design contains molecular docking and *de novo* methods. With the development of genomics and the accumulation of pharmacological information, the big data and deep learning have permeated into the drug discovery fields. For instance, the tensorflow⁵⁵, which is a deep learning software library, has been used for the drug discovery and molecular dynamics simulations⁵⁶. These popular computational methods have been integrated into different software for drug design. The QSAR and pharmacophore model can be constructed by Schrödinger or Discovery Studio software. The molecular docking software can be divided into free academic and commercial programs. Besides, molecular dynamics simulations⁵⁷ are considered as the accurate and dynamical way to study the interaction between targets and ligands. Table 3 shows the free academic, commercial molecular docking programs and molecular dynamics simulations software. LeDock⁵⁸ is designed based on CHARMM force field parameters by using simulated annealing search algorithm. LeDock shows the very high accuracy in pose prediction and is free for the purpose of academic use. rDock is an open source molecular docking program which can be used to dock ligands into the active sites of proteins and nucleic acids⁵⁹. AutoDock Vina and AutoDock are two free academic programs for molecular docking. AutoDock can use the flexibility algorithm to dock the ligands into the proteins by Lamarckian genetic algorithm^{60, 61}. AutoDock Vina, which is considered as the new generation of

AutoDock, has faster run speed and more accurate binding mode predictions than AutoDock⁶². UCSF DOCK⁶³ is the first molecular docking program which contains rigid and flexible ligand docking based on the geometric algorithms⁶⁴. UCSF DOCK can be used to screen the small molecules subset of ZINC database directly. The LigandFit⁶⁵, Glide⁶⁶⁻⁶⁸, GOLD⁶⁹, MOE Dock and Surflex-Dock⁷⁰ use the commercial licenses to service for drug virtual screening⁷¹⁻⁷⁶. They show the powerful ability to screen the drugs from small molecule database through checking their citations on Google scholar. Moreover, wang et al. systematically summarize the advantage on the accuracies of binding pose and binding affinity of molecular docking software by comparing with five free academic license programs (LeDock, rDock, AutoDock, AutoDock Vina, and UCSF DOCK) and five commercial license programs (GOLD, LigandFit, MOE Dock, Glide, and Surflex-Dock)⁷⁷. Furthermore, *de novo* drug design, which is another computer-aided method for drug discovery, can create the new ligands. LigBuilder⁷⁸ and MOE Fragment-Based Design (www.chemcomp.com) are the representative *de novo* drug design software based on fragment linking and growing in the active pocket of targets. The computer-aided software has successfully applied into virtual screening studies and accelerated the process of drug discovery⁷⁹⁻⁸¹. Besides, although the ligands can locate at the active pocket of receptor very well, they may not become the medicine due to the poor ADME (absorption, distribution, metabolism, and excretion)⁸². ADME can be used to build the computer modeling for the prediction of structure-property relationships and reduce the trial failure of drugs in the clinical phases⁸³. The ADME of drugs can be predicted based on the supported molecular format files by Schrödinger or Discovery Studio software.

Molecular docking can perform the high-throughput screening on the huge of small molecule database, while molecular dynamics (MD) simulation is the low-throughput method to evaluate ligand binding pathways⁸⁴. The software Amber⁸⁵, Gromacs⁸⁶, NAMD⁸⁷, and CHARMM⁸⁸ are four popular molecular dynamics package mainly designed for simulations of lipids, nucleic acids and proteins (see Table 3). MD software can be used to study the dynamical interaction between targets and ligands at the atomic level. It can profile more detail and accurate interaction information for

Table 3. Software of molecular docking and molecular dynamics simulations

Software	Description	Weblink
LeDock	Protein-ligand docking	www.lephar.com/software.htm
rDock	Ligands against proteins and nucleic acids	rdock.sourceforge.net
AutoDock	Protein-ligand docking	autodock.scripps.edu
AutoDock Vina	Protein-ligand docking	vina.scripps.edu
UCSF DOCK	Protein-ligand docking	dock.compbio.ucsf.edu
LigandFit	Protein-ligand docking	accelrys.com
Glide	Protein-ligand docking	www.schrodinger.com
GOLD	Protein-ligand docking	www.ccdc.cam.ac.uk/solutions/csd-discovery/components/gold
MOE Dock	Protein-ligand docking	www.chemcomp.com
Surflex-Dock	Protein-ligand docking	www.jainlab.org
Amber	Molecular dynamics simulations	ambermd.org
Gromacs	Molecular dynamics simulations	www.gromacs.org
NAMD	Molecular dynamics simulations	www.ks.uiuc.edu/Research/namd
CHARMM	Molecular dynamics simulations	www.charmm.org

Table 4. Biological assays measuring the binding between macromolecules

Assays	Binding Partners
Surface Plasmon Resonance	Protein-protein, protein-DNA, protein-RNA, protein-drug, antibody-antigen, DNA-DNA
Enzyme-Linked Immunosorbent Assay	Antibody-antigen, protein-ligand
Isothermal Titration Calorimetry	Protein-protein, protein-drug, drug-DNA, protein-DNA, enzyme-substrate
Electrophoretic Mobility Shift Assay	Protein-DNA, protein-RNA
Thermal shift assay	Protein-drug, enzyme-substrate
Protein Fluorescence Quenching	Protein-drug
Differential Scanning Calorimetry	Protein-drug
Nuclear Magnetic Resonance	Protein-drug
Affinity Chromatography	Protein-drug
GST Pull-down	Protein-protein
Footprinting	Protein-DNA
Chromatin Immunoprecipitation	Protein-DNA

ligands in the pocket of receptor than molecular docking. Especially, the Poisson–Boltzmann or generalized Born and surface area continuum solvation (MM/PBSA and MM/GBSA) give the good approaches to compute the binding free energy between ligands and biological macromolecules⁸⁹. As the experiment reported⁹⁰, MM/GBSA shows the faster and better prediction of binding affinities between ligands and targets than MM/PBSA in the absence of metal. MD simulations have become the popular method to study the mechanism of activated, inactivated states and ligand interaction on different targets, such as GPCRs⁹¹⁻⁹⁶, ion channel receptors⁹⁷⁻⁹⁹, etc.

In vitro Biochemical Assays for Compound Screening

One advantage of drug design using high throughput assays and computational tools is that it can largely reduce the use of animals in activity testing. Furthermore, *in vitro* experiments complemented with computational methods have been extensively used in early drug discovery to select compounds with more favorable ADME and toxicological profiles¹⁰⁰⁻¹⁰². Most commonly, drugs are organic small molecules produced through chemical synthesis, but biopolymer-based drugs (also known as biopharmaceuticals) produced through biological processes are becoming increasingly more common. In addition, mRNA-based gene silencing technologies may have therapeutic applications.

The predominant strategy used over the last decades consists, first of all, in clarifying the biochemical processes underlying a disease, then identifying an appropriate drug target and finally developing a suitable assay that allows the screening of chemical libraries for small molecules interfering with the target. Based on the various protein targets of diseases, multiple approaches including biological or *in silico* had been designed to screen new drugs towards the diseases treatment. These approaches are, however, very expensive and demand a great deal of background knowledge. For classical pharmacology, many chemical libraries of synthetic small molecules, natural products or extracts were screened *in vitro* or *in vivo*, such as intact cells, whole organisms or cell-free systems to identify substances that have a desirable therapeutic effect.

The early experimental process to approach drug discovery involves several well defined biochemical

assays to screen those compounds that can interact or bind with certain binding partners, such as receptor/ligand binding analysis^{103, 104}, enzyme-activity evaluation^{105, 106}. In addition, techniques such as X-ray crystallography structural analysis^{107, 108}, NMR^{109, 110}, calorimetry^{111, 112}, affinity chromatography^{113, 114}, ELISA¹¹⁵⁻¹¹⁷ and protein mass spectrometry^{118, 119} are common strategic tools in protein-binding studies and play an important role to enhance the structural basis of rational drug design. These techniques aim to detect the separation of compounds from the studied proteins and monitor changes in intrinsic parameters of the targets upon forming a complex with tested drugs. Table 4 shows the biological assays used to detect the binding event between two binding partners, such as protein-protein, protein-nucleic acid, protein-small molecules. In the next, the applications of isothermal titration calorimetry (ITC) and Surface Plasmon Resonance (SPR) are reviewed in detail.

The rapid development of science technology has prompted the emergence of several new approaches. Isothermal titration calorimetry (ITC) is one of the products of the rapid science technology development. It is most often used to investigate the binding of small molecules to larger macromolecules, such as proteins or DNA. In the measurement using ITC, several important parameters involving the binding process can be calculated, including binding affinity, enthalpy changes, and the binding stoichiometry. According to the obtained parameters, the final Gibbs energy changes and entropy changes can be specifically determined^{120, 121}. As ITC gives not only the binding affinity, but also the thermodynamics of the binding interaction, it is typically used as a secondary screening technique in high throughput drug discovery to eliminate false positive hits after primary screening^{122, 123}. Characterization of the binding thermodynamics allows further hit selection and lead optimization as ITC can provide insights into the structure-activity relationship (SAR) for ligand interaction with the target^{124, 125}. Comparing to other techniques such as fluorescence assays and NMR for studying the complex formation, ITC does not need any fluorescent probes or radioactive tags for data analysis. In addition, proteins used in the measurement do not require chemical modification that is ease of use and cost. In spite of various advantages,

low throughput, low sensitivity, and large sample requirement are major concerns, which may hamper its application¹²⁶⁻¹²⁸.

Another new technique is Surface Plasmon Resonance (SPR), which is the product of nano-science development. The emergence of SPR had greatly reduced the detection limit of biological analysis and it is widely used for the study of ligand binding interactions^{7, 129, 130}. SPR is label-free in that a label molecule is not required for detection of the analytes and capable of measuring real-time quantitative binding affinities and kinetics in sequential binding events^{131, 132}. Moreover, SPR is especially interesting as it can present kinetic information according to affinity data and can be used for thermodynamic studies¹³³⁻¹³⁶. At the same time, SPR biosensor assays can be applied in a wide range of proteins, including membrane proteins, such as G-protein-coupled receptors (GPCRs)¹³⁷⁻¹⁴¹. Another application of SPR technology is early ADME (absorption, distribution, metabolism, and excretion) profile prediction for lead compounds in drug discovery trial¹⁴². SPR have the merits of real-time measurement, label-free and widespread biomolecules, it is emerging as an essential tool for drug development and has been widely used as a primary screening methodology for drug discovery^{125, 143, 144}. During a SPR measurement, the information includes data about concentration of a binding partner in a mixture as well as kinetic rate constants (association, dissociation rate constants and the equilibrium dissociation constant) for the binding interactions¹⁴⁵. Thus, SPR provides insights into the efficacy, safety, duration of action, indication, and patient tolerability of a drug.

Typically, in a compound screening campaign, the selection of evaluation means is highly depended on properties of the target to be studied¹⁴⁶. In studies of prion diseases, researchers have proposed several methods to screen new prion inhibitors that would benefit prion-related patients. As prion protein is prone to convert from a dominantly soluble α -helix structure to β -rich insoluble pathogenic aggregates, efforts have been extensively made toward the aggregation dynamic process and discovery new compounds interrupting the aggregation¹⁴⁷⁻¹⁵⁰. Thioflavin T (ThT) can specifically bind to β -rich aggregates accompanying a red shift of

the fluorescence emission spectra. Hence, it is frequently used as a dye to monitor the aggregation of prion protein¹⁵¹⁻¹⁵⁴. Based on this knowledge, Li et al. evaluated the inhibitory effects of several compounds on prion aggregation using ThT as a detector^{155, 156}. Once the aggregation process is interrupted or interfered by a compound, the increase and intensity of ThT fluorescence are delayed or weakened obviously indicating the efficiency of tested compound. Moreover, by combining multiple approaches in drug screening, it would provide more and much accurate information of the drug-target interaction profile. For example, ITC combined with chromatography has been used to identify and isolate unknown target proteins such as receptors or cell/tissue lysates^{127, 157}. Overall, to conduct a successful drug discovery, the evaluation assays should be carefully selected according to different investigating targets.

In vitro Cell-based Approaches

As a common method used in drug discovery, cell-free systems have various advantages, such as fast, microscale and high throughput and the screening results are accurate and stable^{146, 158, 159}. However, the molecular testing models are designed to specific target, which can only provide limited information about the target interactions. Moreover, the effects of certain drugs on an organism are complex and the interactions between two partners are involved in multiple levels that cannot be predicted using biochemical assays. Hence, single molecular high-throughput drug screening technology can no longer meet the needs of new drug discovery today. Then more biologically relevant cell-based screening assays have been developed and are widely used to predict responses of an organism to drugs^{160, 161}. In addition, the cell culture is selected as a model system to predict cellular toxicity, which plays an important role in drug discovery process¹⁶².

Tumor cell lines are common cell models used in diseases investigation and drug discovery. Current cell-based models rely heavily on immortalized cell lines, usually derived from human tumors. These models have advantages, such as cost-effective scale up and well consistency. Table 5 displays common human-derived cell lines used in cancer-related investigations. For normal cell lines, they are usually used as controls and

Table 5. Common human cell lines used in several cancer researches

Cancer Types	Normal Cell Lines	Cancer Cell Lines
Bladder	SV-HUC-1	T24, 5637, J82
Breast	DU4475, MCF10A	MCF7, SK-BR-3, HCC38,1590
Colon	-	RKO, CW-2, CBZ, SW48, T84, HRC-6, HT-29
Liver	HL-7702, QSG7701, THLE-3, L-02	HepG2, Hep3b, HuH-6, Li-7, PLC/PRF/5, HB611, BEL-7404
Lung	MRC-5, HLF-a, HFL1, WI-38, BEAS-2B	A549, NCI-H157, A427, NCI-H524, TKB-1, Lu-165
Pancreas	HPC-Y5	PANC-1, AsPC-1, HS766T, SW1990
Prostate	WPMY-1, RWPE-1, RWPE-2	DU145, LNCaP, PC-3, 22RV1, VCaP, 2B4
Renal	HEK-293, HKC, 293FT	SW-13, A498, 786-O, Caki-1, 769-P, UT14

toxicology evaluations. Additionally, these cell lines are amenable to genetic engineering, permitting gain and loss of function analysis. While these models demonstrate advantages, they offer limited biological relevance when compared to the intact organ and primary cell types. Currently, primary cells and tissue slices are the gold standards for drug discovery, as they exhibit greater resemblance to the organ of interest¹⁶³⁻¹⁶⁵.

Human pluripotent stem cells hold great promise in research and medicine for their unique ability of self-renewing and differentiating to various cell lineages in the body. For different studies, human pluripotent stem cells can be controlled to differentiate to desired cell types to fulfill the investigation purpose¹⁶⁶⁻¹⁶⁹. As general cell lines used in drug screening present limited relevance to the organ of interest, stem cells provide exciting new models and bring new changes for drug discovery and development as well as drug toxicity testing to treat different human diseases¹⁷⁰. As we known, it is still a great challenge for drug discovery to neurodegenerative diseases because the biological mechanisms are complex and poorly understood. The lack of models that accurately characterize these dysfunctions blocks further investigations. Fortunately, recent advances in stem cell technology offer researchers available tools to generate human neurons

to develop disease resemble assays for small molecules screening. The emergence of adult tissues or cells derived induced pluripotent stem cells (iPSC), which bypass the need for embryos, promotes new investigations of stem cells. Bright et al.¹⁷¹ developed a specific antibody BMS-986168 for the Tau fragment based on human-induced pluripotent stem cells from patients with sporadic Alzheimer's disease (AD). In 2017, this antibody was licensed by Biogen and entered Phase II clinical trials for AD treatment. Retigabine, another drug under clinical Phase II trial was also derived from iPSC models generated from amyotrophic lateral sclerosis (ALS) patients¹⁷². In addition, the recent advances in the production of stem cell-derived hepatocytes and cardiomyocytes combined with cutting-edge engineering technologies supplement the application of stem cells as an attractive alternative model for current drug discovery, which will deliver safer and more efficacious medicines for the patient¹⁷³⁻¹⁷⁵. Moreover, the use of stem cell-derived *in vitro* systems could reduce animal use and facilitate mechanisms investigation of the toxicants at the same time in toxicity evaluations¹⁷⁰. Advancements in pluripotent stem cells and 3D culturing techniques promote the creation of organoids that can accurately recapitulate the properties of various specific subregions of many human organs. Tumor organoids resemble the original tumors much

better than cell lines, having a 3D structure, a variety of cells, and similar growth characteristics, they can be applied as a pre-clinical cancer model for drug discovery^{176, 177}. In addition, organoids provide another opportunity to construct cellular models of human diseases that can be used to deeply study the causes of diseases and further identify possible treatment in laboratory^{178, 179}. We believe that advances in stem cell biology would provide more accurate human tissue and disease models for drug development.

In vivo Animal Models

Compared to *in vitro* screening, *in vivo* testing is better suited for observing the overall effects of an experiment on a living subject. As *in vitro* assays can sometimes yield misleading results with drug candidate molecules that are irrelevant *in vivo*, efficacy verification *in vivo* is especially crucial in drug discovery process. In addition, whole-organism *in vivo* screening holds several advantages to small molecule discovery for its target agonistic and holistic.

One of the most important and widely used model organisms in scientific research is zebrafish, which possesses numerous advantages and is the pioneer model for drug screening. It has been used in various research fields, such as gene expression and sequencing^{180, 181}, cancer models^{182, 183}, immune system¹⁸⁴ and infectious diseases^{185, 186}. Ongoing research programs have promoted zebrafish model to develop novel therapeutic agents in drug discovery. Drug screens based on zebrafish can not only identify novel classes of compounds with biological effects, but also discover novel uses or targets of existing drugs¹⁸⁷. Using a zebrafish screen, a bioactivity from an extract of *Jasminum gilgianum* plant was discovered to induce the formation of ectopic tailbuds in larvae¹⁸⁸. Similarly, some new targets of old medications were identified from zebrafish chemical screening, such as the phenothiazine antipsychotics, which was demonstrated to be toxic to MYC overexpressing thymocytes¹⁸⁹. For the commercially available antiangiogenic statin rosuvastatin, a new function was discovered to suppress the growth of prostate cancer in a zebrafish screening of known bioactive compounds¹⁹⁰. Other new compound classes were also identified based on zebrafish screens, such as GS4012¹⁹¹ and lenalidomide¹⁹².

Another vertebrate animal model frequently used in drug discovery are *Xenopus* frogs, which belong to the amphibians. As *Xenopus* frogs share a long evolutionary history with mammals, they are the excellent models to predict human biology. *Xenopus* have been extensively adopted as a convenient first-line animal model at various stages of drug discovery and development. Since the early 1980s, *Xenopus* embryogenesis has been much explored and a protocol termed frog embryo teratogenesis assay was applied to identify drugs that pose potential teratogenic hazards, including mortality and malformation¹⁹³. Embryos and tadpoles were severed as versatile animal models to investigate blood vascular development and angiogenesis. Subsequently, Roland et al.¹⁹⁴ uncovered pathways involved in the development of the lymphatic and blood vascular system in *Xenopus* tadpoles and discovered new compounds and pathways that were not previously known to mediate lymphatic or vascular development. Due to large number and size of the eggs, the rapid development of the embryos and the fact they are amenable to pharmacological, surgical and genetic techniques, *Xenopus laevis* has been successfully used in searching for embryonic signaling pathways targeting compounds¹⁹⁵, particularly the Wnt/ β -catenin pathway^{196, 197}.

Despite successful application in drug discovery, the zebrafish and *Xenopus* models are largely different from the mammals in various aspects including genetics, immune system, and metabolism. Hence, more advanced mammal models were developed to decrease the gap between animal and human diseases, such as dog, rabbit, rat, mouse and the non-human primate Rhesus and Orangutan Monkey. In a research to determine the isolates of pancreatic secretion could be used to treat dogs with diabetes on an animal model dog, promoted the discovery of insulin and then the use in diabetes treatment¹⁹⁸. For rabbits used as organism models, they are frequently used to produce antibodies in immunology¹⁹⁹. In addition, rabbits are important models to study cardiovascular disease²⁰⁰. Guinea pigs are vertebrate models extensively used by early bacteriologists as hosts for bacterial infections and infectious diseases including viral and parasitic infections²⁰¹. The mouse is one of the classical model vertebrates and has become the popular choice for

developing various *in vivo* mammalian models as it shares about 85% genome identity to humans and has many physiological systems that are similar to those in humans. In addition, mouse has characteristics such as short life-cycle, techniques for genetic manipulation (inbred strains, stem cell lines, and methods of transformation) and non-specialist living requirements that are predominant and convenient to use. It is commonly used for scientific research in medicine, psychology and genetics²⁰². Comparing to the mouse, the rat has larger size of organs and suborganellar structures and it is particularly used as toxicology models and neurological models²⁰³. While for the non-human primate Rhesus and Orangutan Monkeys, they are conventional animal models used in hepatitis, HIV, Parkinson's disease, cognition, and vaccines investigations^{204, 205}. For different scientific research purposes, numerous animal models can be constructed. For example, human tumor xenograft, orthotopic/intratumoral tumor models, murine tumor xenograft and patient-derived tumor grafts were built in nude mice or rats to investigate oncology development or screen new cancer drugs²⁰⁶⁻²⁰⁸. Using the constructed models, development of many human tumors including colon cancer, breast cancer, lung cancer, prostate cancer, ovarian cancer, renal cancer, cervical cancer, pancreatic cancer and melanoma has been investigated at different degrees. At the same time, therapeutic compounds targeting these tumors were also discovered either from the existing drugs or from new synthetic compounds.

Despite numerous models developed for drug discovery, most therapeutic drugs still fail in clinical trials. One of the reasons is attributed to sufficient clinical predictive power of our current model systems. Despite the high genetic similarities between human and mice, physiological differences affect the course of diseases in mice models when some genetic disorders in human do not have the same symptoms in mice. In addition, the cell lines and xenografts commonly used are inadequate models that can not highly mimic and accurately predict human diseases. Generally, for a drug discovery research in the laboratory, in combination with the chemical assays, cell-based and *in vivo* testing would perform more efficiently to obtain effective lead compounds for further drug development. At last, new models are still needed to be developed for scientific

researches in the future.

Conclusions

As above review, the entire drug design process can be profiled from *in silico*, *in vitro* and *in vivo* experiments (see Figure 1). Our review summarizes the functions of macromolecular structure databases, small molecule databases, molecular docking software, *de novo* drug design software, MD simulations software, biochemical assays, cell-based system and animal models. This review shows the detailed individual component of drug design, and gives the comprehensive understanding for the progress of drug design.

Acknowledgments

The work is supported by the National Natural Science Foundation of China (Grant No. 21605066) and Fundamental Research Funds for the Central Universities (Grant No. lzujbky-2018-92).

Additional information

Competing financial interests: The authors declare no competing financial interests.

References

1. Sertkaya, A., Wong, H. H., Jessup, A., & Beleche, T. (2016) Key cost drivers of pharmaceutical clinical trials in the United States. *Clin. Trials* 13, 117-126.
2. Kim, R. S., Goossens, N., & Hoshida, Y. (2016) Use of big data in drug development for precision medicine. *Expert Rev Precis Med Drug Dev* 1, 245-253.
3. Pagadala, N. S., Syed, K., & Tuszynski, J. (2017) Software for molecular docking: a review. *Biophys Rev* 9, 91-102.
4. Schneider, G. & Fechner, U. (2005) Computer-based *de novo* design of drug-like molecules. *Nat. Rev. Drug Discov.* 4, 649-663.
5. Liu, X., Shi, D., Zhou, S., Liu, H., Liu, H., *et al.* (2018) Molecular dynamics simulations and novel drug discovery. *Expert Opin Drug Discov* 13, 23-37.
6. Basith, S., Lee, Y., & Choi, S. (2018) Understanding G Protein-Coupled Receptor Allostery via Molecular Dynamics Simulations: Implications for Drug Discovery. *Methods Mol. Biol.* 1762, 455-472.
7. Zeng, S., Baillargeat, D., Ho, H. P., & Yong, K. T. (2014) Nanomaterials enhanced surface plasmon

- resonance for biological and chemical sensing applications. *Chem. Soc. Rev.* 43, 3426-3452.
8. Mazur, M., Bujak, A., Matloka, M., Janowska, S., Gunerka, P., *et al.* (2015) Cell-based assay for low- and high-scale screening of the Wnt/beta-catenin signaling modulators. *Anal. Biochem.* 475, 56-67.
 9. Daher, A. & de Groot, J. (2018) Rapid identification and validation of novel targeted approaches for Glioblastoma: A combined ex vivo-in vivo pharmacomic model. *Exp. Neurol.* 299, 281-288.
 10. Bai, Q. (2018) Big Data Research: Database and Computing. *Journal of Big Data Research* 1, 1-4.
 11. Berman, H., Henrick, K., & Nakamura, H. (2003) Announcing the worldwide Protein Data Bank. *Nat. Struct. Biol.* 10, 980.
 12. Berman, H. M., Westbrook, J., Feng, Z., Gilliland, G., Bhat, T. N., *et al.* (2000) The Protein Data Bank. *Nucleic Acids Res.* 28, 235-242.
 13. Wuthrich, K. (2001) The way to NMR structures of proteins. *Nat. Struct. Biol.* 8, 923-925.
 14. Callaway, E. (2015) The Revolution Will Not Be Crystallized. *Nature* 525, 172-174.
 15. Coimbatore Narayanan, B., Westbrook, J., Ghosh, S., Petrov, A. I., Sweeney, B., *et al.* (2014) The Nucleic Acid Database: new features and capabilities. *Nucleic Acids Res.* 42, D114-122.
 16. Madej, T., Address, K. J., Fong, J. H., Geer, L. Y., Geer, R. C., *et al.* (2012) MMDB: 3D structures and macromolecular interactions. *Nucleic Acids Res.* 40, D461-464.
 17. Reichert, J. & Suhnel, J. (2002) The IMB Jena Image Library of Biological Macromolecules: 2002 update. *Nucleic Acids Res.* 30, 253-254.
 18. Wang, R., Fang, X., Lu, Y., Yang, C. Y., & Wang, S. (2005) The PDBbind database: methodologies and updates. *J. Med. Chem.* 48, 4111-4119.
 19. Wang, R., Fang, X., Lu, Y., & Wang, S. (2004) The PDBbind database: collection of binding affinities for protein-ligand complexes with known three-dimensional structures. *J. Med. Chem.* 47, 2977-2980.
 20. Siddiquee, K., Zhang, S., Guida, W. C., Blaskovich, M. A., Greedy, B., *et al.* (2007) Selective chemical probe inhibitor of Stat3, identified through structure-based virtual screening, induces antitumor activity. *Proc. Natl. Acad. Sci. U. S. A.* 104, 7391-7396.
 21. Cross, J. B. (2018) Methods for Virtual Screening of GPCR Targets: Approaches and Challenges. *Methods Mol. Biol.* 1705, 233-264.
 22. Bai, Q., Shao, Y., Pan, D., Zhang, Y., Liu, H., *et al.* (2014) Search for beta2 adrenergic receptor ligands by virtual screening via grid computing and investigation of binding modes by docking and molecular dynamics simulations. *PLoS One* 9, e107837.
 23. Bai, Q. & Yao, X. (2016) Investigation of allosteric modulation mechanism of metabotropic glutamate receptor 1 by molecular dynamics simulations, free energy and weak interaction analysis. *Sci. Rep.* 6, 21763.
 24. Djikic, T., Marti, Y., Spyrikis, F., Lau, T., Benedetti, P., *et al.* (2018) Human dopamine transporter: the first implementation of a combined in silico/in vitro approach revealing the substrate and inhibitor specificities. *J Biomol Struct Dyn*, 1-16.
 25. Kristam, R., Rao, S. N., D'Cruz, A. S., Mahadevan, V., & Viswanadhan, V. N. (2017) TRPV1 antagonism by piperazinyl-aryl compounds: A Topomer-CoMFA study and its use in virtual screening for identification of novel antagonists. *J. Mol. Graph. Model.* 72, 112-128.
 26. Alexander, S. P., Kelly, E., Marrion, N. V., Peters, J. A., Faccenda, E., *et al.* (2017) THE CONCISE GUIDE TO PHARMACOLOGY 2017/18: Overview. *Br J Pharmacol* 174 Suppl 1, S1-S16.
 27. Harding, S. D., Sharman, J. L., Faccenda, E., Southan, C., Pawson, A. J., *et al.* (2018) The IUPHAR/BPS Guide to PHARMACOLOGY in 2018: updates and expansion to encompass the new guide to IMMUNOPHARMACOLOGY. *Nucleic Acids Res* 46, D1091-D1106.
 28. Pieper, U., Webb, B. M., Dong, G. Q., Schneidman-Duhovny, D., Fan, H., *et al.* (2014) ModBase, a database of annotated comparative protein structure

- models and associated resources. *Nucleic Acids Res.* 42, D336-346.
29. Haas, J., Roth, S., Arnold, K., Kiefer, F., Schmidt, T., *et al.* (2013) The Protein Model Portal--a comprehensive resource for protein structure and model information. *Database (Oxford)* 2013, bat031.
30. Biasini, M., Bienert, S., Waterhouse, A., Arnold, K., Studer, G., *et al.* (2014) SWISS-MODEL: modelling protein tertiary and quaternary structure using evolutionary information. *Nucleic Acids Res.* 42, W252-W258.
31. Leffler, A. E., Kuryatov, A., Zebroski, H. A., Powell, S. R., Filipenko, P., *et al.* (2017) Discovery of peptide ligands through docking and virtual screening at nicotinic acetylcholine receptor homology models. *Proc. Natl. Acad. Sci. U. S. A.* 114, E8100-E8109.
32. Feng, Z., Pearce, L. V., Xu, X., Yang, X., Yang, P., *et al.* (2015) Structural insight into tetrameric hTRPV1 from homology modeling, molecular docking, molecular dynamics simulation, virtual screening, and bioassay validations. *J. Chem. Inf. Model.* 55, 572-588.
33. Irwin, J. J., Sterling, T., Mysinger, M. M., Bolstad, E. S., & Coleman, R. G. (2012) ZINC: a free tool to discover chemistry for biology. *J. Chem. Inf. Model.* 52, 1757-1768.
34. Irwin, J. J. & Shoichet, B. K. (2005) ZINC--a free database of commercially available compounds for virtual screening. *J. Chem. Inf. Model.* 45, 177-182.
35. Modi, P., Patel, S., & Chhabria, M. T. (2018) Identification of some novel pyrazolo[1,5-a]pyrimidine derivatives as InhA inhibitors through pharmacophore-based virtual screening and molecular docking. *J. Biomol Struct Dyn*, 1-14.
36. Zhang, J., Zhu, N., Du, Y., Bai, Q., Chen, X., *et al.* (2015) Dehydrocrenatidine is a novel janus kinase inhibitor. *Mol. Pharmacol.* 87, 572-581.
37. Kim, S., Thiessen, P. A., Bolton, E. E., Chen, J., Fu, G., *et al.* (2016) PubChem Substance and Compound databases. *Nucleic Acids Res.* 44, D1202-1213.
38. Wishart, D. S., Knox, C., Guo, A. C., Cheng, D., Shrivastava, S., *et al.* (2008) DrugBank: a knowledgebase for drugs, drug actions and drug targets. *Nucleic Acids Res.* 36, D901-906.
39. Dietrich, R. C., Alberca, L. N., Ruiz, M. D., Palestro, P. H., Carrillo, C., *et al.* (2018) Identification of cisapride as new inhibitor of putrescine uptake in *Trypanosoma cruzi* by combined ligand- and structure-based virtual screening. *Eur. J. Med. Chem.* 149, 22-29.
40. Van Den Driessche, G. & Fourches, D. (2018) Adverse drug reactions triggered by the common HLA-B*57:01 variant: virtual screening of DrugBank using 3D molecular docking. *J. Cheminform.* 10, 3.
41. Park, I., Londhe, A. M., Lim, J. W., Park, B. G., Jung, S. Y., *et al.* (2017) Discovery of non-peptidic small molecule inhibitors of cyclophilin D as neuroprotective agents in Abeta-induced mitochondrial dysfunction. *J. Comput. Aided Mol. Des.* 31, 929-941.
42. Fu, Y., Sun, Y. N., Yi, K. H., Li, M. Q., Cao, H. F., *et al.* (2017) 3D Pharmacophore-Based Virtual Screening and Docking Approaches toward the Discovery of Novel HPPD Inhibitors. *Molecules* 22.
43. Park, I., Hwang, Y. J., Kim, T., Viswanath, A. N. I., Londhe, A. M., *et al.* (2017) In silico probing and biological evaluation of SETDB1/ESET-targeted novel compounds that reduce tri-methylated histone H3K9 (H3K9me3) level. *J. Comput. Aided Mol. Des.* 31, 877-889.
44. Pihan, E., Colliandre, L., Guichou, J. F., & Douguet, D. (2012) e-Drug3D: 3D structure collections dedicated to drug repurposing and fragment-based drug design. *Bioinformatics* 28, 1540-1541.
45. Dunkel, M., Fullbeck, M., Neumann, S., & Preissner, R. (2006) SuperNatural: a searchable database of available natural compounds. *Nucleic Acids Res.* 34, D678-683.
46. Mohamed, A., Nguyen, C. H., & Mamitsuka, H. (2016) Current status and prospects of computational resources for natural product dereplication: a review. *Brief Bioinform* 17, 309-321.
47. Groom, C. R., Bruno, I. J., Lightfoot, M. P., & Ward, S. C. (2016) The Cambridge Structural Database.

- Acta Crystallogr B Struct Sci Cryst Eng Mater 72, 171-179.
48. Grazulis, S., Daskevicius, A., Merkys, A., Chateigner, D., Lutterotti, L., *et al.* (2012) Crystallography Open Database (COD): an open-access collection of crystal structures and platform for world-wide collaboration. *Nucleic Acids Res.* 40, D420-427.
49. Goto, S., Okuno, Y., Hattori, M., Nishioka, T., & Kanehisa, M. (2002) LIGAND: database of chemical compounds and reactions in biological pathways. *Nucleic Acids Res.* 30, 402-404.
50. Kumar Mishra, S. & Kumar, A. (2016) NALDB: nucleic acid ligand database for small molecules targeting nucleic acid. *Database (Oxford)* 2016.
51. Jewison, T., Su, Y., Disfany, F. M., Liang, Y., Knox, C., *et al.* (2014) SMPDB 2.0: big improvements to the Small Molecule Pathway Database. *Nucleic Acids Res.* 42, D478-484.
52. Prathipati, P., Dixit, A., & Saxena, A. K. (2007) Computer-aided drug design: Integration of structure-based and ligand-based approaches in drug design. *Current Computer-Aided Drug Design* 3, 133-148.
53. Yousefinejad, S. & Hemmateenejad, B. (2015) Chemometrics tools in QSAR/QSPR studies: A historical perspective. *Chemometrics and Intelligent Laboratory Systems* 149, 177-204.
54. Wermuth, C., Ganellin, C., Lindberg, P., & Mitscher, L. (1998) Glossary of terms used in medicinal chemistry (IUPAC Recommendations 1998). *Pure Appl. Chem.* 70, 1129-1143.
55. Abadi, M., Barham, P., Chen, J., Chen, Z., Davis, A., *et al.* (2016) Tensorflow: a system for large-scale machine learning. *OSDI*, pp 265-283.
56. Yao, K., Herr, J. E., Toth, D. W., Mckintyre, R., & Parkhill, J. (2018) The TensorMol-0.1 model chemistry: a neural network augmented with long-range physics. *Chemical science* 9, 2261-2269.
57. Alder, B. J. & Wainwright, T. E. (1959) Studies in Molecular Dynamics. I. General Method. *The Journal of Chemical Physics* 31, 459-466.
58. Zhang, N. & Zhao, H. (2016) Enriching screening libraries with bioactive fragment space. *Bioorg. Med. Chem. Lett.* 26, 3594-3597.
59. Ruiz-Carmona, S., Alvarez-Garcia, D., Foloppe, N., Garmendia-Doval, A. B., Juhos, S., *et al.* (2014) rDock: a fast, versatile and open source program for docking ligands to proteins and nucleic acids. *PLoS Comput. Biol.* 10, e1003571.
60. Morris, G. M., Goodsell, D. S., Halliday, R. S., Huey, R., Hart, W. E., *et al.* (1998) Automated docking using a Lamarckian genetic algorithm and an empirical binding free energy function. *J. Comput. Chem.* 19, 1639-1662.
61. Morris, G. M., Huey, R., Lindstrom, W., Sanner, M. F., Belew, R. K., *et al.* (2009) AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J. Comput. Chem.* 30, 2785-2791.
62. Trott, O. & Olson, A. J. (2010) AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *J. Comput. Chem.* 31, 455-461.
63. Lang, P. T., Brozell, S. R., Mukherjee, S., Pettersen, E. F., Meng, E. C., *et al.* (2009) DOCK 6: combining techniques to model RNA-small molecule complexes. *RNA* 15, 1219-1230.
64. Kuntz, I. D., Blaney, J. M., Oatley, S. J., Langridge, R., & Ferrin, T. E. (1982) A geometric approach to macromolecule-ligand interactions. *J. Mol. Biol.* 161, 269-288.
65. Venkatachalam, C. M., Jiang, X., Oldfield, T., & Waldman, M. (2003) LigandFit: a novel method for the shape-directed rapid docking of ligands to protein active sites. *J Mol Graph Model* 21, 289-307.
66. Friesner, R. A., Murphy, R. B., Repasky, M. P., Frye, L. L., Greenwood, J. R., *et al.* (2006) Extra precision glide: docking and scoring incorporating a model of hydrophobic enclosure for protein-ligand complexes. *J. Med. Chem.* 49, 6177-6196.
67. Halgren, T. A., Murphy, R. B., Friesner, R. A., Beard, H. S., Frye, L. L., *et al.* (2004) Glide: a new approach for rapid, accurate docking and scoring. 2. Enrichment factors in database screening. *J. Med. Chem.* 47, 1750-1759.

68. Friesner, R. A., Banks, J. L., Murphy, R. B., Halgren, T. A., Klicic, J. J., *et al.* (2004) Glide: a new approach for rapid, accurate docking and scoring. 1. Method and assessment of docking accuracy. *J. Med. Chem.* 47, 1739-1749.
69. Jones, G., Willett, P., Glen, R. C., Leach, A. R., & Taylor, R. (1997) Development and validation of a genetic algorithm for flexible docking. *J. Mol. Biol.* 267, 727-748.
70. Jain, A. N. (2003) Surflex: Fully automatic flexible molecular docking using a molecular similarity-based search engine. *J. Med. Chem.* 46, 499-511.
71. Lather, A., Sharma, S., & Khatkar, A. (2018) Virtual Screening of Novel Glucosamine-6-Phosphate Synthase Inhibitors. *Comb. Chem. High Throughput Screen.* 21, 182-193.
72. Larif, S., Salem, C. B., Hmouda, H., & Bouraoui, K. (2014) In silico screening and study of novel ERK2 inhibitors using 3D QSAR, docking and molecular dynamics. *J. Mol. Graph. Model.* 53, 1-12.
73. Doytchinova, I., Atanasova, M., Valkova, I., Stavrov, G., Philipova, I., *et al.* (2018) Novel hits for acetylcholinesterase inhibition derived by docking-based screening on ZINC database. *J. Enzyme Inhib. Med. Chem.* 33, 768-776.
74. Wang, N., Wang, L., & Xie, X. Q. (2017) ProSelection: A Novel Algorithm to Select Proper Protein Structure Subsets for in Silico Target Identification and Drug Discovery Research. *J. Chem. Inf. Model.* 57, 2686-2698.
75. Wadood, A., Ghufuran, M., Hassan, S. F., Khan, H., Azam, S. S., *et al.* (2017) In silico identification of promiscuous scaffolds as potential inhibitors of 1-deoxy-d-xylulose 5-phosphate reductoisomerase for treatment of Falciparum malaria. *Pharm. Biol.* 55, 19-32.
76. Bai, Q., Zhang, Y., Jin, L., & Yao, X. (2018) Applying Virtual Drug Screening Workflow Module of Schrödinger into College Bioinformatics and Chemoinformatics Teaching. *University Chemistry* 33, 66-71.
77. Wang, Z., Sun, H. Y., Yao, X. J., Li, D., Xu, L., *et al.* (2016) Comprehensive evaluation of ten docking programs on a diverse set of protein-ligand complexes: the prediction accuracy of sampling power and scoring power. *PCCP* 18, 12964-12975.
78. Yuan, Y. X., Pei, J. F., & Lai, L. H. (2011) LigBuilder 2: A Practical de Novo Drug Design Approach. *J. Chem. Inf. Model.* 51, 1083-1091.
79. Amaro, R. E., Baudry, J., Chodera, J., Demir, O., McCammon, J. A., *et al.* (2018) Ensemble Docking in Drug Discovery. *Biophys. J.*
80. Vilar, S., Sobarzo-Sanchez, E., Santana, L., & Uriarte, E. (2017) Molecular Docking and Drug Discovery in beta-Adrenergic Receptors. *Curr. Med. Chem.* 24, 4340-4359.
81. Nan, J., Du, Y., Chen, X., Bai, Q., Wang, Y., *et al.* (2014) TPCA-1 is a direct dual inhibitor of STAT3 and NF-kappaB and regresses mutant EGFR-associated human non-small cell lung cancers. *Mol. Cancer Ther.* 13, 617-629.
82. Tremaine, L., Brian, W., DelMonte, T., Francke, S., Groenen, P., *et al.* (2015) The role of ADME pharmacogenomics in early clinical trials: perspective of the Industry Pharmacogenomics Working Group (I-PWG). *Pharmacogenomics* 16, 2055-2067.
83. Lombardo, F., Desai, P. V., Arimoto, R., Desino, K. E., Fischer, H., *et al.* (2017) In Silico Absorption, Distribution, Metabolism, Excretion, and Pharmacokinetics (ADME-PK): Utility and Best Practices. An Industry Perspective from the International Consortium for Innovation through Quality in Pharmaceutical Development. *J Med Chem* 60, 9097-9113.
84. Sledz, P. & Caflisch, A. (2018) Protein structure-based drug design: from docking to molecular dynamics. *Curr. Opin. Struct. Biol.* 48, 93-102.
85. Salomon-Ferrer, R., Case, D. A., & Walker, R. C. (2013) An overview of the Amber biomolecular simulation package. *Wiley Interdisciplinary Reviews-Computational Molecular Science* 3, 198-210.
86. Van Der Spoel, D., Lindahl, E., Hess, B., Groenhof, G., Mark, A. E., *et al.* (2005) GROMACS: fast, flexible, and free. *J. Comput. Chem.* 26, 1701-1718.

87. Phillips, J. C., Braun, R., Wang, W., Gumbart, J., Tajkhorshid, E., *et al.* (2005) Scalable molecular dynamics with NAMD. *J Comput Chem* 26, 1781-1802.
88. Brooks, B. R., Brooks, C. L., 3rd, Mackerell, A. D., Jr., Nilsson, L., Petrella, R. J., *et al.* (2009) CHARMM: the biomolecular simulation program. *J Comput Chem* 30, 1545-1614.
89. Genheden, S. & Ryde, U. (2015) The MM/PBSA and MM/GBSA methods to estimate ligand-binding affinities. *Expert Opin Drug Discov* 10, 449-461.
90. Hou, T., Wang, J., Li, Y., & Wang, W. (2011) Assessing the performance of the MM/PBSA and MM/GBSA methods. 1. The accuracy of binding free energy calculations based on molecular dynamics simulations. *J. Chem. Inf. Model.* 51, 69-82.
91. Bai, Q., Zhang, Y., Li, X., Chen, W., Liu, H., *et al.* (2014) Computational study on the interaction between CCR5 and HIV-1 entry inhibitor maraviroc: insight from accelerated molecular dynamics simulation and free energy calculation. *Phys. Chem. Chem. Phys.* 16, 24332-24338.
92. Bai, Q., Perez-Sanchez, H., Zhang, Y., Shao, Y., Shi, D., *et al.* (2014) Ligand induced change of beta2 adrenergic receptor from active to inactive conformation and its implication for the closed/open state of the water channel: insight from molecular dynamics simulation, free energy calculation and Markov state model analysis. *Phys. Chem. Chem. Phys.* 16, 15874-15885.
93. Bai, Q., Shi, D., Zhang, Y., Liu, H., & Yao, X. (2014) Exploration of the antagonist CP-376395 escape pathway for the corticotropin-releasing factor receptor 1 by random acceleration molecular dynamics simulations. *Mol. Biosyst.* 10, 1958-1967.
94. Bai, Q., Shen, Y., Jin, N., Liu, H., & Yao, X. (2014) Molecular modeling study on the dynamical structural features of human smoothed receptor and binding mechanism of antagonist LY2940680 by metadynamics simulation and free energy calculation. *Biochim. Biophys. Acta* 1840, 2128-2138.
95. Bai, Q., Zhang, Y., Ban, Y., Liu, H., & Yao, X. (2013) Computational study on the different ligands induced conformation change of beta2 adrenergic receptor-Gs protein complex. *PLoS One* 8, e68138.
96. Bai, Q., Perez-Sanchez, H., Shi, Z., Li, L., Shi, D., *et al.* (2018) Computational studies on horseshoe shape pocket of human orexin receptor type 2 and boat conformation of suvorexant by molecular dynamics simulations. *Chem Biol Drug Des* 92, 1221-1231.
97. Li, G. H. (2018) Exploring conformational states and helical packings in the P2X receptor transmembrane domain by molecular dynamics simulation. *J Biol Phys.*
98. Martin, N. E., Malik, S., Calimet, N., Changeux, J. P., & Cecchini, M. (2017) Un-gating and allosteric modulation of a pentameric ligand-gated ion channel captured by molecular dynamics. *PLoS Comput Biol* 13, e1005784.
99. Harpole, T. J. & Delemotte, L. (2018) Conformational landscapes of membrane proteins delineated by enhanced sampling molecular dynamics simulations. *Biochim Biophys Acta* 1860, 909-926.
100. Ripphausen, P., Nisius, B., Peltason, L., & Bajorath, J. (2010) Quo vadis, virtual screening? A comprehensive survey of prospective applications. *J Med Chem* 53, 8461-8467.
101. Clark, D. E. (2008) What has virtual screening ever done for drug discovery? *Expert Opin Drug Discov* 3, 841-851.
102. McGregor, M. J., Luo, Z., & Jiang, X. (2007) *Chapter 3: Virtual screening in drug discovery* (Wiley-VCH: Weinheim, Germany) p 25.
103. Hulme, E. C. & Trevethick, M. A. (2010) Ligand binding assays at equilibrium: validation and interpretation. *British journal of pharmacology* 161, 1219-1237.
104. Pollard, T. D. (2010) A guide to simple and informative binding assays. *Molecular biology of the cell* 21, 4061-4067.
105. Goddard, J.-P. & Reymond, J.-L. (2004) Enzyme assays for high-throughput screening. *Current opinion in biotechnology* 15, 314-322.
106. Acker, M. G. & Auld, D. S. (2014) Considerations for the design and reporting of enzyme assays in

- high-throughput screening applications. *Perspectives in Science* 1, 56-73.
107. Davis, A. M., Teague, S. J., & Kleywegt, G. J. (2003) Application and limitations of X-ray crystallographic data in structure-based ligand and drug design. *Angew. Chem. Int. Ed. Engl.* 42, 2718-2736.
108. Blundell, T. L., Jhoti, H., & Abell, C. (2002) High-throughput crystallography for lead discovery in drug design. *Nat. Rev. Drug Discov.* 1, 45-54.
109. Roberts, G. C. (2000) Applications of NMR in drug discovery. *Drug discovery today* 5, 230-240.
110. Pellecchia, M., Sem, D. S., & Wuthrich, K. (2002) NMR in drug discovery. *Nat. Rev. Drug Discov.* 1, 211-219.
111. Weber, P. C. & Salemme, F. R. (2003) Applications of calorimetric methods to drug discovery and the study of protein interactions. *Curr. Opin. Struct. Biol.* 13, 115-121.
112. Henriques, D. A. & Ladbury, J. E. (2001) Inhibitors to the Src SH2 domain: a lesson in structure--thermodynamic correlation in drug design. *Arch. Biochem. Biophys.* 390, 158-168.
113. Ascoli, G. A., Domenici, E., & Bertucci, C. (2006) Drug binding to human serum albumin: Abridged review of results obtained with high-performance liquid chromatography and circular dichroism. *Chirality* 18, 667-679.
114. Kim, H. S. & Wainer, I. W. (2008) Rapid analysis of the interactions between drugs and human serum albumin (HSA) using high-performance affinity chromatography (HPAC). *Journal of chromatography. B, Analytical technologies in the biomedical and life sciences* 870, 22-26.
115. Lequin, R. M. (2005) Enzyme immunoassay (EIA)/enzyme-linked immunosorbent assay (ELISA). *Clinical chemistry* 51, 2415-2418.
116. Alonso, N., Guillen, R., Chambers, J. W., & Leng, F. (2015) A rapid and sensitive high-throughput screening method to identify compounds targeting protein-nucleic acids interactions. *Nucleic acids research* 43, e52.
117. Fung, E., Sugianto, P., Hsu, J., Damoiseaux, R., Ganz, T., *et al.* (2013) High-throughput screening of small molecules identifies hepcidin antagonists. *Molecular pharmacology* 83, 681-690.
118. Glish, G. L. & Vachet, R. W. (2003) The basics of mass spectrometry in the twenty-first century. *Nature reviews. Drug discovery* 2, 140-150.
119. de Mol, N. J., Catalina, M. I., Fischer, M. J., Broutin, I., Maier, C. S., *et al.* (2004) Changes in structural dynamics of the Grb2 adaptor protein upon binding of phosphotyrosine ligand to its SH2 domain. *Biochimica et biophysica acta* 1700, 53-64.
120. Pierce, M. M., Raman, C. S., & Nall, B. T. (1999) Isothermal Titration Calorimetry of Protein-Protein Interactions. *Methods* 19, 213-221.
121. Lai, M. & Lau, Y. (2018) Measurement of binding strength between prey proteins interacting with *Toxoplasma gondii* SAG1 and SAG2 using isothermal titration calorimetry (ITC). *Acta Parasitol.* 63, 106-113.
122. Peters, W. B., Frasca, V., & Brown, R. K. (2009) Recent developments in isothermal titration calorimetry label free screening. *Combinatorial chemistry & high throughput screening* 12, 772-790.
123. Leavitt, S. & Freire, E. (2001) Direct measurement of protein binding energetics by isothermal titration calorimetry. *Current opinion in structural biology* 11, 560-566.
124. Sarver, R. W., Peevers, J., Cody, W. L., Ciske, F. L., Dyer, J., *et al.* (2007) Binding thermodynamics of substituted diaminopyrimidine renin inhibitors. *Anal. Biochem.* 360, 30-40.
125. Ferenczy, G. G. & Keseru, G. M. (2010) Thermodynamics guided lead discovery and optimization. *Drug Discov. Today* 15, 919-932.
126. Liang, Y. (2008) Applications of isothermal titration calorimetry in protein science. *Acta Biochim Biophys Sin (Shanghai)* 40, 565-576.
127. Zhou, X., Sun, Q., Kini, R. M., & Sivaraman, J. (2008) A universal method for fishing target proteins from mixtures of biomolecules using isothermal titration calorimetry. *Protein science : a publication of the Protein Society* 17, 1798-1804.
128. Ladbury, J. E., Klebe, G., & Freire, E. (2010) Adding calorimetric data to decision making in lead

- discovery: a hot tip. *Nature reviews. Drug discovery* 9, 23-27.
129. Tang, Y., Zeng, X., & Liang, J. (2010) Surface Plasmon Resonance: An Introduction to a Surface Spectroscopy Technique. *Journal of chemical education* 87, 742-746.
130. Patching, S. G. (2014) Surface plasmon resonance spectroscopy for characterisation of membrane protein–ligand interactions and its potential for drug discovery. *Biochimica et Biophysica Acta (BBA) - Biomembranes* 1838, 43-55.
131. Rich, R. L. & Myszka, D. G. (2007) Higher-throughput, label-free, real-time molecular interaction analysis. *Analytical Biochemistry* 361, 1-6.
132. Saha, K., Agasti, S. S., Kim, C., Li, X., & Rotello, V. M. (2012) Gold nanoparticles in chemical and biological sensing. *Chem. Rev.* 112, 2739-2779.
133. Shuman, C. F., Markgren, P. O., Hamalainen, M., & Danielson, U. H. (2003) Elucidation of HIV-1 protease resistance by characterization of interaction kinetics between inhibitors and enzyme variants. *Antiviral research* 58, 235-242.
134. Gabrielsson, J., Dolgos, H., Gillberg, P. G., Bredberg, U., Benthem, B., *et al.* (2009) Early integration of pharmacokinetic and dynamic reasoning is essential for optimal development of lead compounds: strategic considerations. *Drug discovery today* 14, 358-372.
135. Markgren, P. O., Schaal, W., Hamalainen, M., Karlen, A., Hallberg, A., *et al.* (2002) Relationships between structure and interaction kinetics for HIV-1 protease inhibitors. *Journal of medicinal chemistry* 45, 5430-5439.
136. Katsamba, P. S., Park, S., & Laird-Offringa, I. A. (2002) Kinetic studies of RNA-protein interactions using surface plasmon resonance. *Methods* 26, 95-104.
137. Navratilova, I. & Hopkins, A. L. (2011) Emerging role of surface plasmon resonance in fragment-based drug discovery. *Future medicinal chemistry* 3, 1809-1820.
138. Lukong, K. E., Chang, K. W., Khandjian, E. W., & Richard, S. (2008) RNA-binding proteins in human genetic disease. *Trends in genetics : TIG* 24, 416-425.
139. Walhout, A. J. (2006) Unraveling transcription regulatory networks by protein-DNA and protein-protein interaction mapping. *Genome research* 16, 1445-1454.
140. Rich, R. L. & Myszka, D. G. (2003) A survey of the year 2002 commercial optical biosensor literature. *Journal of molecular recognition : JMR* 16, 351-382.
141. Khan, S. H., Farkas, K., Kumar, R., & Ling, J. (2012) A versatile method to measure the binding to basic proteins by surface plasmon resonance. *Anal Biochem* 421, 385-390.
142. Fabini, E. & Danielson, U. H. (2017) Monitoring drug-serum protein interactions for early ADME prediction through Surface Plasmon Resonance technology. *Journal of pharmaceutical and biomedical analysis* 144, 188-194.
143. Hajduk, P. J. & Greer, J. (2007) A decade of fragment-based drug design: strategic advances and lessons learned. *Nature reviews. Drug discovery* 6, 211-219.
144. Huber, W. (2005) A new strategy for improved secondary screening and lead optimization using high - resolution SPR characterization of compound - target interactions. *Journal of Molecular Recognition* 18, 273-281.
145. Minunni M., B. A. R. (2010) SPR in Drug Discovery: Searching Bioactive Compounds in Plant Extracts. *Ligand-Macromolecular Interactions in Drug Discovery*, (Humana Press, Totowa, NJ, Totowa), Vol 572, pp 203-218.
146. Fara, D. C., Oprea, T. I., Prossnitz, E. R., Bologa, C. G., Edwards, B. S., *et al.* (2006) Integration of virtual and physical screening. *Drug Discovery Today: Technologies* 3, 377-385.
147. Bamdad, K. & Naderi-Manesh, H. (2007) Contribution of a putative salt bridge and backbone dynamics in the structural instability of human prion protein upon R208H mutation. *Biochem Biophys Res Commun* 364, 719-724.

148. Behmard, E., Abdolmaleki, P., Asadabadi, E. B., & Jahandideh, S. (2011) Prevalent mutations of human prion protein: a molecular modeling and molecular dynamics study. *J Biomol Struct Dyn* 29, 379-389.
149. Chen, W., van der Kamp, M. W., & Daggett, V. (2010) Diverse Effects on the Native beta-Sheet of the Human Prion Protein Due to Disease-Associated Mutations. *Biochemistry* 49, 9874-9881.
150. Rossetti, G., Giachin, G., Legname, G., & Carloni, P. (2010) Structural facets of disease-linked human prion protein mutants: a molecular dynamic study. *Proteins* 78, 3270-3280.
151. Biancalana, M. & Koide, S. (2010) Molecular mechanism of Thioflavin-T binding to amyloid fibrils. *Biochimica et biophysica acta* 1804, 1405-1412.
152. Rolinski, O. J., Amaro, M., & Birch, D. J. (2010) Early detection of amyloid aggregation using intrinsic fluorescence. *Biosens Bioelectron* 25, 2249-2252.
153. Zhou, Z., Yan, X., Pan, K., Chen, J., Xie, Z. S., *et al.* (2011) Fibril formation of the rabbit/human/bovine prion proteins. *Biophys J* 101, 1483-1492.
154. Zhou, Z., Fan, J. B., Zhu, H. L., Shewmaker, F., Yan, X., *et al.* (2009) Crowded cell-like environment accelerates the nucleation step of amyloidogenic protein misfolding. *J Biol Chem* 284, 30148-30158.
155. Li, L., Wei, W., Jia, W. J., Zhu, Y., Zhang, Y., *et al.* (2017) Discovery of small molecules binding to the normal conformation of prion by combining virtual screening and multiple biological activity evaluation methods. *J Comput Aided Mol Des* 31, 1053-1062.
156. Li, L., Zhu, Y., Zhou, S., An, X., Zhang, Y., *et al.* (2017) Experimental and Theoretical Insights into the Inhibition Mechanism of Prion Fibrillation by Resveratrol and its Derivatives. *ACS Chem. Neurosci.* 8, 2698-2707.
157. Zhou, X., Kini, R. M., & Sivaraman, J. (2011) Application of isothermal titration calorimetry and column chromatography for identification of biomolecular targets. *Nature protocols* 6, 158-165.
158. Martis, E. A. R., R.; Badve, R.R. (2011) High-throughput screening: The hits and leads of drug discovery—An overview. *Journal of Applied Pharmaceutical Science* 1, 2-10.
159. Fernandes, T. G., Diogo, M. M., Clark, D. S., Dordick, J. S., & Cabral, J. M. (2009) High-throughput cellular microarray platforms: applications in drug discovery, toxicology and stem cell research. *Trends in biotechnology* 27, 342-349.
160. Sundberg, S. A. (2000) High-throughput and ultra-high-throughput screening: solution- and cell-based approaches. *Current opinion in biotechnology* 11, 47-53.
161. Ciambrone, G. J., Liu, V. F., Lin, D. C., McGuinness, R. P., Leung, G. K., *et al.* (2004) Cellular dielectric spectroscopy: a powerful new approach to label-free cellular analysis. *Journal of biomolecular screening* 9, 467-480.
162. Szymanski, P., Markowicz, M., & Mikiciuk-Olasik, E. (2012) Adaptation of high-throughput screening in drug discovery-toxicological screening tests. *International journal of molecular sciences* 13, 427-452.
163. Horvath, P., Aulner, N., Bickle, M., Davies, A. M., Nery, E. D., *et al.* (2016) Screening out irrelevant cell-based models of disease. *Nature reviews. Drug discovery* 15, 751-769.
164. Masters, J. R. & Stacey, G. N. (2007) Changing medium and passaging cell lines. *Nature protocols* 2, 2276-2284.
165. Nestor, C. E., Ottaviano, R., Reinhardt, D., Cruickshanks, H. A., Mjoseng, H. K., *et al.* (2015) Rapid reprogramming of epigenetic and transcriptional profiles in mammalian culture systems. *Genome biology* 16, 11.
166. Takahashi, K. & Yamanaka, S. (2006) Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 126, 663-676.
167. Selvaraj, V., Plane, J. M., Williams, A. J., & Deng, W. (2010) Switching cell fate: the remarkable rise of induced pluripotent stem cells and lineage reprogramming technologies. *Trends in biotechnology* 28, 214-223.

- 168.Selvaraj, V., Jiang, P., Chechneva, O., Lo, U. G., & Deng, W. (2012) Differentiating human stem cells into neurons and glial cells for neural repair. *Frontiers in bioscience* 17, 65-89.
- 169.Bai, Q., Zhang, Y., Jin, L., & Yao, X. (2018) Applying Virtual Drug Screening Workflow Module of Schrödinger into College Bioinformatics and Chemoinformatics Teaching. *University Chemistry* 33.
- 170.Liu, W., Deng, Y., Liu, Y., Gong, W., & Deng, W. (2013) Stem cell models for drug discovery and toxicology studies. *Journal of biochemical and molecular toxicology* 27, 17-27.
- 171.Bright, J., Hussain, S., Dang, V., Wright, S., Cooper, B., *et al.* (2015) Human secreted tau increases amyloid-beta production. *Neurobiol. Aging* 36, 693-709.
- 172.Wainger, B. J., Kiskinis, E., Mellin, C., Wiskow, O., Han, S. S., *et al.* (2014) Intrinsic membrane hyperexcitability of amyotrophic lateral sclerosis patient-derived motor neurons. *Cell Rep* 7, 1-11.
- 173.Donato, M. T., Jover, R., & Gomez-Lechon, M. J. (2013) Hepatic cell lines for drug hepatotoxicity testing: limitations and strategies to upgrade their metabolic competence by gene engineering. *Current drug metabolism* 14, 946-968.
- 174.Pradip, A., Steel, D., Jacobsson, S., Holmgren, G., Ingelman-Sundberg, M., *et al.* (2016) High Content Analysis of Human Pluripotent Stem Cell Derived Hepatocytes Reveals Drug Induced Steatosis and Phospholipidosis. *Stem cells international* 2016, 2475631.
- 175.Takayama, K. & Mizuguchi, H. (2017) Generation of human pluripotent stem cell-derived hepatocyte-like cells for drug toxicity screening. *Drug metabolism and pharmacokinetics* 32, 12-20.
- 176.Weeber, F., Ooft, S. N., Dijkstra, K. K., & Voest, E. E. (2017) Tumor Organoids as a Pre-clinical Cancer Model for Drug Discovery. *Cell Chemical Biology* 24, 1092-1100.
- 177.van de Wetering, M., Francies, Hayley E., Francis, Joshua M., Bounova, G., Iorio, F., *et al.* (2015) Prospective Derivation of a Living Organoid Biobank of Colorectal Cancer Patients. *Cell* 161, 933-945.
- 178.Nakano, T., Ando, S., Takata, N., Kawada, M., Muguruma, K., *et al.* (2012) Self-formation of optic cups and storable stratified neural retina from human ESCs. *Cell stem cell* 10, 771-785.
- 179.Huch, M. & Koo, B.-K. (2015) Modeling mouse and human development using organoid cultures. *Development* 142, 3113-3125.
- 180.Kimmel, C. B. & Law, R. D. (1985) Cell lineage of zebrafish blastomeres. I. Cleavage pattern and cytoplasmic bridges between cells. *Dev. Biol.* 108, 78-85.
- 181.Kimmel, C. B. & Law, R. D. (1985) Cell lineage of zebrafish blastomeres. III. Clonal analyses of the blastula and gastrula stages. *Dev. Biol.* 108, 94-101.
- 182.Liu, S. & Leach, S. D. (2011) Zebrafish models for cancer. *Annu. Rev. Pathol.* 6, 71-93.
- 183.White, R. M., Cech, J., Ratanasirintrao, S., Lin, C. Y., Rahl, P. B., *et al.* (2011) DHODH modulates transcriptional elongation in the neural crest and melanoma. *Nature* 471, 518-522.
- 184.Le Guyader, D., Redd, M. J., Colucci-Guyon, E., Murayama, E., Kissa, K., *et al.* (2008) Origins and unconventional behavior of neutrophils in developing zebrafish. *Blood* 111, 132-141.
- 185.Meecker, N. D. & Trede, N. S. (2008) Immunology and zebrafish: spawning new models of human disease. *Dev. Comp. Immunol.* 32, 745-757.
- 186.Renshaw, S. A. & Trede, N. S. (2012) A model 450 million years in the making: zebrafish and vertebrate immunity. *Dis. Model. Mech.* 5, 38-47.
- 187.Rennekamp, A. J. & Peterson, R. T. (2015) 15 years of zebrafish chemical screening. *Curr. Opin. Chem. Biol.* 24, 58-70.
- 188.Gebruers, E., Cordero-Maldonado, M. L., Gray, A. I., Clements, C., Harvey, A. L., *et al.* (2013) A phenotypic screen in zebrafish identifies a novel small-molecule inducer of ectopic tail formation suggestive of alterations in non-canonical Wnt/PCP signaling. *PLoS One* 8, e83293.

189. Gutierrez, A., Pan, L., Groen, R. W., Baleyrier, F., Kentsis, A., *et al.* (2014) Phenothiazines induce PP2A-mediated apoptosis in T cell acute lymphoblastic leukemia. *J. Clin. Invest.* 124, 644-655.
190. Wang, C., Tao, W., Wang, Y., Bikow, J., Lu, B., *et al.* (2010) Rosuvastatin, identified from a zebrafish chemical genetic screen for antiangiogenic compounds, suppresses the growth of prostate cancer. *Eur. Urol.* 58, 418-426.
191. Peterson, R. T., Shaw, S. Y., Peterson, T. A., Milan, D. J., Zhong, T. P., *et al.* (2004) Chemical suppression of a genetic mutation in a zebrafish model of aortic coarctation. *Nat. Biotechnol.* 22, 595-599.
192. Ridges, S., Heaton, W. L., Joshi, D., Choi, H., Eiring, A., *et al.* (2012) Zebrafish screen identifies novel compound with selective toxicity against leukemia. *Blood* 119, 5621-5631.
193. Dumont, J. N., Schultz, T. W., Buchanan, M. V., & Kao, G. L. (1983) Frog Embryo Teratogenesis Assay: Xenopus (FETAX) — A Short-Term Assay Applicable to Complex Environmental Mixtures. *Short-Term Bioassays in the Analysis of Complex Environmental Mixtures III*, eds Waters MD, Sandhu SS, Lewtas J, Claxton L, Chernoff N, & Nesnow S (Springer US, Boston, MA), pp 393-405.
194. Kalin, R. E., Banziger-Tobler, N. E., Detmar, M., & Brandli, A. W. (2009) An in vivo chemical library screen in Xenopus tadpoles reveals novel pathways involved in angiogenesis and lymphangiogenesis. *Blood* 114, 1110-1122.
195. Maia, L. A., Velloso, I., & Abreu, J. G. (2017) Advances in the use of Xenopus for successful drug screening. *Expert Opin Drug Discov* 12, 1153-1159.
196. Nathalia, G. A., Barbara, F. F., Debora Malta, C., Alice, H. R., Alessandro Bolis Costa, S., *et al.* (2012) Effects of Natural Compounds on Xenopus Embryogenesis: A Potential Read Out for Functional Drug Discovery Targeting Wnt/β-catenin Signaling. *Current Topics in Medicinal Chemistry* 12, 2103-2113.
197. Dominguez, I. & Green, J. B. (2000) Dorsal downregulation of GSK3β by a non-Wnt-like mechanism is an early molecular consequence of cortical rotation in early Xenopus embryos. *Development* 127, 861-868.
198. Karamitsos, D. T. (2011) The story of insulin discovery. *Diabetes research and clinical practice* 93, S2-S8.
199. Esteves, P. J. d. C. (2003) Molecular and population genetic analysis of polymorphism at the antibody loci IgGCH2 and IgVH in lagomorphs.
200. Pogwizd, S. M. & Bers, D. M. (2008) Rabbit models of heart disease. *Drug Discovery Today: Disease Models* 5, 185-193.
201. Padilla-Carlin, D. J., McMurray, D. N., & Hickey, A. J. (2008) The guinea pig as a model of infectious diseases. *Comparative medicine* 58, 324-340.
202. Piret, S. E. & Thakker, R. V. (2017) Mouse Models: Approaches to Generate In Vivo Models for Hereditary Disorders of Mineral and Skeletal Homeostasis. *Genetics of Bone Biology and Skeletal Disease (Second Edition)*, (Elsevier), pp 89-118.
203. Aikawa, H., Nonaka, I., Woo, M., Tsugane, T., & Esaki, K. (1988) Shaking rat Kawasaki (SRK): a new neurological mutant rat in the Wistar strain. *Acta neuropathologica* 76, 366-372.
204. Molina, P. E., Amedee, A. M., Winsauer, P., Nelson, S., Bagby, G., *et al.* (2015) Behavioral, metabolic, and immune consequences of chronic alcohol or cannabinoids on HIV/AIDS: studies in the non-human primate SIV model. *Journal of Neuroimmune Pharmacology* 10, 217-232.
205. Zhou, Q. (2014) Balancing the welfare: the use of non-human primates in research. *Trends in Genetics* 30, 476-478.
206. Siolas, D. & Hannon, G. J. (2013) Patient-derived tumor xenografts: transforming clinical samples into mouse models. *Cancer research* 73, 5315-5319.
207. Morton, C. L. & Houghton, P. J. (2007) Establishment of human tumor xenografts in immunodeficient mice. *Nature protocols* 2, 247-250.
208. Richmond, A. & Su, Y. (2008) Mouse xenograft models vs GEM models for human cancer therapeutics. *Disease models & mechanisms* 1, 78-82.